

**ANTIOXIDANT CAPACITY IN ORGANICALLY AND  
CONVENTIONALLY GROWN MANGO (*MANGIFERA INDICA* L.)  
AND PINEAPPLE (*ANANAS COSMUS*)**

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**ABSTRACT:** *Organically grown mango (Mangifera indica L. cv. Dodo, Bolibo, Viringe) and pineapple (Ananas comosus L. cv. Sooth cayenne) from Tanzania were compared to organically (cv Tommy Atkins) and conventionally (cv Kent) grown mango and pineapple (cv Smooth cayenne) cultivars from seven other countries purchased in markets in Germany. The influence of cultivar, agricultural practice and geographical location on total antioxidant capacity, total phenolics, carotenoids and ascorbic acid levels was determined. Cultivars, agricultural practices and geographical location have shown influence on some of the quality attributes of the fruit. Ascorbic acid was significantly higher in conventional than in organically produced mango and pineapple at different locations. Antioxidant capacity was higher by 22% in conventionally than organically grown mango. The total carotenoids content was 17% higher in organic than in conventional mango, and 21% higher in conventional than in organically produced pineapple.*

**KEYWORDS:** Total antioxidant capacity, total phenolic compounds, total carotenoids and ascorbic acid, mango, pineapple

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## **INTRODUCTION**

Organic food sales have increased by about 20% per year since 1990, and were estimated at \$10.4 billion in 2003 on the U.S. market alone (Oberholtz et al., 2005). Consumer studies have shown multiple reasons for organic preferences, including environmental and socioeconomic concerns, enhanced taste, and the assumption that organic foods are healthier (Shepherd et al 2005).

Quality values distinguish between species depending on genetically-determined differences. The quality target for crop production is to reach the best possible composition of compounds or groups of compounds that are important in nutrition and consumer health. These compounds are created in primary and secondary synthesis pathways in plants (Lundegårdh, 2005).

Agronomic practices play a role in determining the quality of food with respect to health and safety attributes, differing soil management between organic or conventional production are key determinants of differences in quality and safety. Very few studies have compared nutrient and bioactive compounds in organic and conventionally produced foods (Lombardi-Boccia et al., 2004).

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High consumption of fruits and vegetables has been associated with a lower incidence of degenerative diseases in consumers (Ribeiro et al., 2007). Such protective effects are thought to be partially associated with the presence of various antioxidant compounds (Kauer and Kapoor, 2001). Mango (*Mangifera indica* L.) and pineapple (*Ananas cosmos*) are among the most important commercial crops in Tanzania. These fruits are considered as a good source of dietary antioxidants, such as amino acids, carotenoids and phenolic compounds (Sánchez-Moreno et al., 2006).

Fruit and vegetables contain various bioactive compounds such as vitamins A, C and E, with a high antioxidant capacity (Hassimoto et al., 2005 and Sánchez-Moreno et al., 2006). and phenolic compounds, which are good contributors to the total antioxidant capacity of the foods (Cano et al., 2003 and Chaovanalikit and Wrolstad, 2004), although their nutritional relevance is uncertain because they may be poorly absorbed and rapidly metabolized and thus have limited antioxidant ability *in vivo* (Gardner et al., 2000).

The study of natural antioxidants has become popular due to increased demand in the market for so-called functional or health foods (Andlauer and Fürst, 2002). Ascorbic acid (AA) is an essential human diet component and acts as an antioxidant and therefore offers some protection against oxidative stress-related diseases (Zulueta et al., 2007). The action of phenolic compounds in foods has received some attention because of their biological activity in cancer and heart diseases prevention (Kris-Etherton et al 2002). These compounds are preferably oxidized in biological medium and function as antioxidant nutrient economizer, protecting organisms against the oxidative stress (Ribeiro et al., 2008).

Among the carotenoid pigments widely distributed in plant tissues,  $\beta$ -carotene provides the highest vitamin A activity. Vitamin A and its metabolites are essential for vision, reproduction, and immune function, besides performing other important physiological functions, including the deactivation of reactive oxygen species (Ball, 2004). Fruit contains various carotenoids, which are a group of red, orange, and yellow pigments. Several of the carotenoids are vitamin A precursors. Carotenoids also function as quenchers of singlet oxygen, as antioxidants, in gene activation, and in inflammation and immune processes as a modulator of lipoxygenases (Ajila et al., 2007). There is a significant correlation between the high intake of fruit and vegetables and the low incidence of some kinds of cancer and cardiovascular disease (Dragsted et al., 2006). This may be due to the protective effect of fruit and vegetables which are caused by the presence of a wide range of different antioxidants found in dietary plants (Halvorsenet al., 2002). While antioxidants are produced by the human body as well, the antibody defence mechanism could be further strengthened with a diet rich in antioxidant compounds. Several antioxidants with varied chemical characteristics are found to act together in a synergistic way (Blomhoff et al., 2006). A wide range of antioxidants was found in dietary plants as potential sources especially fruits, vegetables and nuts as well (Halvorsen et al., 2002).

The aim of this study was to compare the influence of agricultural practice, cultivar and geographical location on antioxidant capacity, phenolic content, carotenoids content and ascorbic acid levels of mango and pineapple fruit

## **MATERIALS AND METHODS**

### **Sampling**

Organically grown mango (*Mangifera indiga* L. cv. *Dodo*, *Bolibo*, *Viringe*) and pineapple (*Ananas comosus* L. cv. *Sooth cayenne*) from Tanzania were obtained from local markets in Dar

es Salaam, Tanzania and transported by flight to Germany. Fruits obtained from shops in Göttingen, Germany were organic mango *cv. Tommy Atkins* from Burkina Faso, conventional mango *cv. Kent* from Ivory Coast, Mali, Peru and Costa Rica, organic pineapple *cv. Smooth cayenne* from Uganda and conventional pineapple *cv. Smooth cayenne* from Ghana and Honduras. All fruits used were obtained between January and March in 2006, 2007 and 2008 and analysed. All fruits were at acceptable ripeness stage and their eating quality was considered. They were grouped into five fruits each for mango and pineapple. Fruits were cooled to 5-7 °C before cutting.

### **Ferric reducing antioxidant power (FRAP method)**

The antioxidant capacity of fruits was estimated using the FRAP assay (Connor *et al.*, 2002), using a spectrophotometer (Hewlett Packard 8453, Germany).

The antioxidant capacity of a fruit was determined by its ability to reduce ferric iron to ferrous in a solution of 2, 4, 6-tripyridyl-2-triazine (TPTZ) prepared in sodium acetate at pH 3.6. The reduction of iron in the TPTZ-ferric chloride solution (FRAP reagent) results in the formation of a blue product (ferrous tripyridyltriazine complex). The FRAP assay was done by putting 100µl of the sample into the cuvette and 1 ml FRAP reagent was added and mixed thoroughly, then incubated for 4 min at 37°C in a water bath, the absorbance was measured at 593 nm against zero blank (1 ml FRAP reagent and 100 µl methanol). The antioxidant standard curve (50 -1000 mM ferrous ion) was developed using ferrous ammonium sulphate. The results are expressed as millimoles of ferrous equivalents per gram of fresh weight.

### **Total phenolic content**

The total phenolic substances were determined photometrically using the Folin-Ciocalteu method (Singleton and Rossi, 1965)

The calibration curve was prepared with different concentrations of gallic acid (the same amount of a 0.5 mol L<sup>-1</sup> sodium hydroxide solution and Folin-Ciocalteu reagent). For the measurement 500 µl of sample, 1 ml of sodium hydroxide solution, 100 µl Folin-Ciocalteu reagent and 2.4 ml distilled water were used, mixed and set into the water bath (T=37°C) and incubated for 15 minutes. The measurements were done on a spectrophotometer (Hewlett Packard 8453, Germany) at 735.8nm.

The total phenol contents of fruit juices were determined in triplicate and expressed in gallic acid equivalents (GAE).

### **Total carotenoids content**

According to (Wellburn, 1994) 0.2 g of freeze dried sample was put in 5 ml of methanol and mixed on Vortex for one minute, then was shaken for 10 minutes and finally centrifuged at 3500 rev/min for 15 minutes. The extraction with methanol was repeated three times and finally the supernatants were measured at 470, 563 and 666 nm absorbance with spectrophotometer (Hewlett Packard 8453, Germany). The following equation was used for total carotenoid determination.

Chlorophyll a (Chl a)	$(12.19 \times A_{665}) - (3.45 \times A_{649})$	µg/ml
Chlorophyll b (Chl b)	$(21.19 \times A_{649}) - (5.32 \times A_{665})$	µg/ml
Total Carotenoid	$((1000 \times A_{480}) - (2.14 \times \text{Chl a}) - (70.16 \times \text{Chl b})) / 220$	µg/ml

### **Ascorbic acid**

AA was determined by titration method (Albrecht, 1993). Five grams samples were immersed in 20 ml metaphosphoric acid (5%) in 50 ml tube and homogenized by ultra turrax for 2 minutes

and after the addition of distilled water up to the volume. Then the solution was filtered using filter paper. The filtrate (10 ml) was titrated with 0.21 % 2, 6 dichlorophenol-indiphenol (DIP) solution drop by drop until light red orange colour was achieved. The amount of DIP solution used was recorded for the calculation of the AA content in the sample tested using the formula: Ascorbic acid (mg100g<sup>-1</sup>juice) = ml DIP x 0.1 x (50/10) x (100/weight sample)

### Statistical analysis

Collected data were entered into a spreadsheet and imported into SigmaStat program version 2, where various statistical tests were performed using ANOVA. Least significant different (LSD) test, a multiple comparison test, was performed in an attempt to discern if significant differences existed between any of the sample means.

## RESULTS AND DISCUSSION

### Antioxidant capacity in fruits

Fruits and vegetables are rich sources of several micronutrients and phytochemicals with antioxidant properties, which are said to be protective against chronic degenerative health disorders (Cox et al., 2000 and Szeto et al., 2002) These compounds with antioxidant properties include vitamin C, vitamin E, carotenoids and phenolic compounds, especially flavonoids. Fruits and vegetables have antioxidants which could act against oxidative stress, protect cells against oxidative damage and also prevent chronic diseases like cancer, cardiovascular disease and diabetes (Podsedek, 2005). Normally, nutrient antioxidants function as synergists, due to this they are more effective together rather than as single antioxidant in lowering the levels of oxygen species (Eberhardt et al., 2000; Ohr, 2004) Trombino et al., 2004; Podsedek, 2005). Fruits and vegetables contain other important nutrients which are essential to human health because the human body is not able to produce them. These substances include water, protein, lipid, minerals and /or vitamins.

Table 1.1: Antioxidant capacity in mango and pineapple fruits

Fruit	Antioxidant capacity (mMol Fe <sup>2+</sup> 100g <sup>-1</sup> FW)	Origin	Production
Mango cultivars			
Dodo	0.48±0.18abc	Tanzania	Organic
Bolibo	0.45±0.17bc	Tanzania	Organic
Viringe	0.42±0.08c	Tanzania	Organic
Tommy Atkins	0.36±0.06c	Burkina Faso	Organic
Kent	0.49±0.1abc	Costa Rica	Conventional
Kent	0.63±0.17a	Ivory Coast	Conventional
Kent	0.61±0.13ab	Mali	Conventional
Kent	0.49±0.04abc	Peru	Conventional
Pineapple cultivar			
Smooth cayenne	0.34±0.04b	Tanzania	Organic
Smooth cayenne	0.57±0.15a	Uganda	Organic
Smooth cayenne	0.56±0.29a	Costa Rica	Conventional
Smooth cayenne	0.44±0.2ab	Ghana	Conventional
Smooth cayenne	0.45±0.14ab	Honduras	Conventional

Values within columns of the same fruits with different letters are significantly different ( $p \leq 0.05$ ).

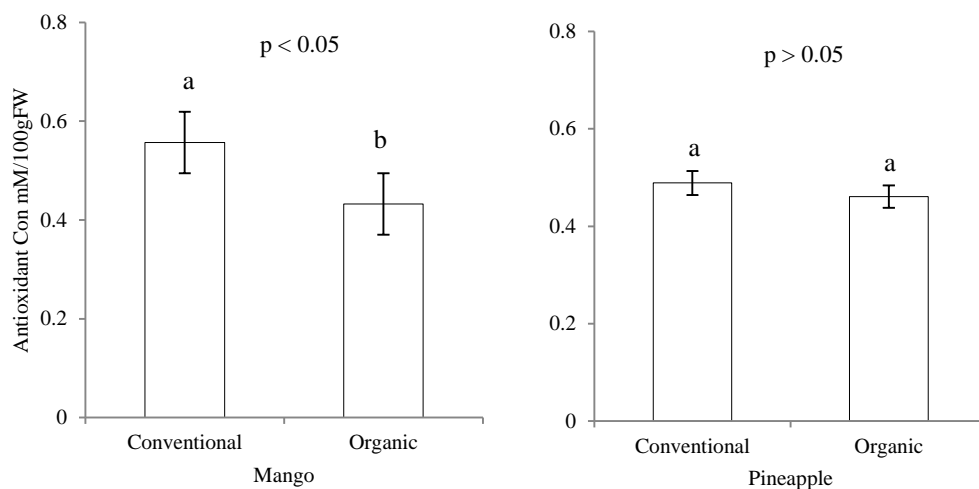


Figure 1.1: Total antioxidant capacity in conventional and organic in mango (n=12) and pineapple fruits (n=16). Values with different letters are significantly different ( $p \leq 0.05$ ).

In Table 2.1 the total antioxidant capacities have shown variations within the mango cultivars, which were significant at  $p \leq 0.05$ . Mango cultivar 'Kent' have shown the highest and 'Tommy Atkins' the lowest value in their total antioxidant capacity, which could be influenced by cultivar differences. The mango cultivar 'Kent' produced under conventional production system in different locations differed significantly in their total antioxidant capacities. This suggests that the geographical location may influence the total antioxidant of the mango cultivar due to different soil types and climate conditions. The total antioxidant capacities of mango cultivars under conventional production systems were 22% higher compared with mango cultivars under organic production system (Fig. 2.1), showing that the production systems could affect the total antioxidant capacity of mango fruits.

The pineapple cultivar 'Smooth Cayenne' from different countries has shown significant differences in their total antioxidant capacities ( $p \leq 0.05$ ), which implies that there is effect of location on total antioxidant capacities. The conventional pineapple fruits have shown 5.7% higher total antioxidant capacity than organically produced pineapple, indicating the influence of cultivation methods on the levels of the total antioxidant capacity in pineapple fruits. (Young et al., 2005 and Chassy et al., 2006) found that different crops may respond differently to agronomic factors such as nutrient availability. There is evidence suggesting that higher levels of antioxidant and vitamins in organically than those conventionally produced (Weibel et al., 2000; Asami et al., 2003 and Chassy et al., 2006) However, there is also a study that shows either results that are opposite, or results that show no difference (Barrett et al., 2007). Winter and Davis, (2006) concluded that it is not possible to ensure that, from a nutritional point of view, organically grown products are superior to those obtained by conventional agricultural techniques.

### Total carotenoids content of fruits

In a study done by Rao and Rao (2007), it was found that fruits and vegetables are major source of carotenoids. These compounds contain conjugated double bonds which cause them to act as antioxidants (Sandmann, 2001). Recently, further studies have been done on carotenoids effects on antioxidant capacity and the ability to prevent diseases. Carotenoids are used as food additives (Breithaupt, 2004) and the yellow-reddish colour of many foods are due to the

presence of carotenoids, which are natural pigments found abundantly in fruits and vegetables (Dutta et al, 2005 and Mortensen, 2006). According to Rodriguez-Amaya (2001) mango fruits could provide provitamin A carotenoids, especially  $\beta$ -carotene.

Table 1.2: Total carotenoids content in mango and pineapple fruit

Fruit	Total carotenoids (mg 100g <sup>-1</sup> FW)	Origin	Production
Mango cultivars			
Dodo	2.13±0.82ab	Tanzania	Organic
Bolibo	1.87±0.68ab	Tanzania	Organic
Viringe	2.87±0.97a	Tanzania	Organic
Tommy Atkins	1.52±0.89b	Burkina Faso	Organic
Kent	1.96±0.75ab	Costa Rica	Conventional
Kent	1.37±0.55b	Ivory Coast	Conventional
Kent	2.21±1.06ab	Mali	Conventional
Kent	1.76±0.95ab	Peru	Conventional
Pineapple cultivar			
Smooth cayenne	1.17±0.46a	Tanzania	Organic
Smooth cayenne	1.98±0.32a	Uganda	Organic
Smooth cayenne	2.0±1.0a	Costa Rica	Conventional
Smooth cayenne	1.69±1.0a	Ghana	Conventional
Smooth cayenne	1.58±1.0a	Honduras	Conventional

Values within columns of the same fruits with different letters are significantly different ( $p \leq 0.05$ ).

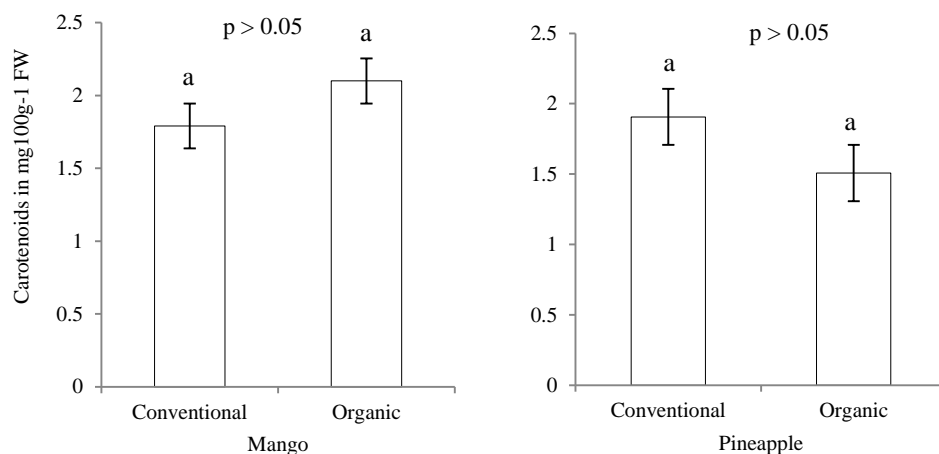


Figure 1.2: Total carotenoid content in mango (n=12) and pineapple fruits (n=16). Values with same letters are not significantly different ( $p \geq 0.05$ ).

The mango cultivars showed significant differences at ( $p \leq 0.05$ ) in their total content of carotenoids (Table 2.2). Mango cultivar 'Viringe' have shown highest content in the total carotenoids content of 2.87mg 100g<sup>-1</sup> FW. The mango cultivars 'Kent' grown at different locations varied significantly in their total carotenoid contents, indicating that location may influence the total carotenoid contents of the fruits. The total carotenoid content was 17.3% higher in mango cultivars from organic than in conventional production systems (Fig. 2.2). This may imply that the agricultural methods could have effect on total carotenoids content of the fruits. (Winter and Davis, 2006) stated that fruits and vegetables crops grown in organic farms are not applied with chemical pesticides or synthetic fertilizers and instead they synthesize beneficial secondary plant metabolites such as polyphenolic antioxidants as well as naturally

occurring toxins. Climatic effects are seen in fruits of the same cultivars cultivated in regions of different climates, elevated temperature and greater exposure to sunlight, thus increasing carotenogenesis. The total carotenoids contents were found to range from 1.17 to 2.06 mg 100g<sup>-1</sup>FW in pineapple fruits cultivar '*Smooth Cayenne*'. (Joomwong, 2006) found the pineapple cultivar '*Smooth Cayenne*' to have average carotenoid content between 1 to 1.5 mg 100g<sup>-1</sup>FW. In our study pineapple cultivar '*Smooth Cayenne*' has shown slight differences which were not significant ( $p \geq 0.05$ ) in their total carotenoid content. Stracke et al. (2009) found in their study that carotenoids content did not vary between organic and conventional grown fruits. Although, the total carotenoid was not significant ( $p \geq 0.05$ ) among pineapple cultivars, the total carotenoid content was on average 21% higher in pineapple fruits under conventional than organic production system (Fig. 2.2), which suggests that cultivation system may influence on carotenoid content of the pineapple fruits. Due to rapidly available nitrogen in the conventional farming system plant diverts sugars from photosynthesis to produce more proteins and a spike in vegetative growth. As a result plant produces more leaves, and thus more chloroplasts, which will bring out higher carotenoids production. Rodriguez-Amaya (2001) stated that carotenoid content in fruits could be influenced by some differences such as the stage of maturity, cultivar, and post-harvest handling procedures. There are qualitative differences due to factors such as cultivar or variety, climate or geographic site of production and farming practices, which could influence carotenoids contents in food (Dutta et al. (2005).

### **Total phenolics content of fruits**

Phenolic compounds belong to secondary metabolites which are by-products of physiological processes occurring in plants. Studies done by Podsedek, (2005) and Balasundram et al., (2006) found these derivatives of pentose phosphate, shikimate and phenylpropanoid pathways have shown a strong antioxidant effect against free radicals as well as to other reactive oxygen species. Natural phenolic antioxidants have shown health promoting benefits and, because of this, fruits and vegetables became more important in human nutrition. The bio-functional health-promoting properties of phenolic compounds led scientists to investigate their availability in fruits and vegetable, as important sources of such compound (Moure et al., 2001). Health promoting properties of fruits were recently being associated with phenolic compounds, since natural occurring antioxidants have shown to contribute against oxidative damage caused by free radicals. (Lattanzio, 2003) stated that phenolic compounds shown health promoting properties of fruits including beneficial influence also against disease development such as cancer and coronary heart diseases. Useniket al. (2004) found that the level of susceptibility/tolerance to fungal infections and pests could be determined by phenolic composition of plant tissue.

Table 1.3: Total phenolic content in mango and pineapple fruits

Fruit	Total phenolic content (mg100 ml <sup>-1</sup> juice)	Origin	Production
Mango cultivars			
Dodo	2.71±0.44ab	Tanzania	Organic
Bolibo	3.08±0.17a	Tanzania	Organic
Viringe	2.77±0.40ab	Tanzania	Organic
Tommy Atkins	2.47±0.11b	Burkina Faso	Organic
Kent	2.69±0.64ab	Costa Rica	Conventional
Kent	2.99±0.20a	Ivory Coast	Conventional
Kent	2.72±0.33ab	Mali	Conventional
Kent	3.07±0.05a	Peru	Conventional
Pineapple cultivar			
Smooth cayenne	2.95±0.11a	Tanzania	Organic
Smooth cayenne	3.05±0.43a	Uganda	Organic
Smooth cayenne	2.99±0.21a	Costa Rica	Conventional
Smooth cayenne	2.91±0.38a	Ghana	Conventional
Smooth cayenne	2.93±0.36a	Honduras	Conventional

Value within columns of the same fruits with different letters is significantly different ( $p \leq 0.05$ ).

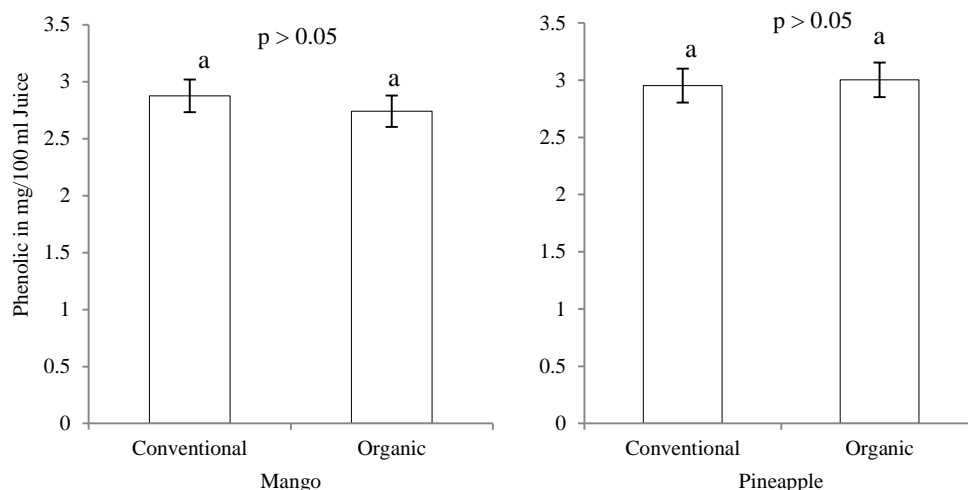


Figure 1.3: Total phenolic content in mango (n=12) and pineapple fruits (n=16). Values with same letters are not significantly different ( $p \geq 0.05$ ).

The total phenol contents of fruit juices were expressed as gallic acid equivalents (GAE) using the Folin-Ciocalteu method. The total phenolic content cultivars vary significantly at  $p \leq 0.05$  among mango cultivars (Table 2.3), and it was on average 4.7% higher in conventional than in organic mango fruits. (Cartea et al., 2011) found in their study that maturity stage, environmental conditions, especially light, and mode of fertilization could influence the production of phenolic compounds. Mango cultivars 'Bolibo' and 'Kent' have shown highest value in their total phenolic compounds (3.08 and 3.06 mg100 ml<sup>-1</sup> juice). The variation of total phenolic content between mango cultivars could be due the genetic variations, as it is well-known that the biosynthesis of phenolic compounds in plants is strongly influenced by the cultivar (Häkkinen and Törrönen, 2000).



The difference in total phenolic content was not significant ( $p \geq 0.05$ ) among pineapple fruits (Table 2.3), although organic pineapples have shown 1.7% higher than conventional pineapples in their phenolic contents (Fig. 2.3). Anttonen et al. (2006) observed that the variations of fruit phenolics contents were not significant between organic and conventional fruits grown in farms within a similar climatic conditions. Valavanidis et al. (2009) found in their study that in polyphenol content was similar for both organic and conventionally grown fruits. Other study done by Veberic and Stampar, (2005) found that apple grown organically have shown higher content of phenolic compounds in the pulp compared to conventional apples. Carbonaro et al. (2002) revealed that phenolic substances increase in organic peaches and pear fruit production. This speculates the role of phenolic compound in defence mechanism of plant, due to the fact that in organic agriculture where there is no application of chemical pesticides, plants use their natural compounds to defend themselves against pest and disease. The method of cultivation and location could contribute to the difference in the total phenolic content of pineapple cultivar 'smooth cayenne', although, in our study the effect of conventional and organic production system has shown slight influence. This could be due to different soil types and climate condition which these pineapple cultivars were produced. The soils differ in their availability of nutrients to plant due to different agricultural practices which could be influenced by the method of fertilization. In organic agriculture, the nitrogen is realised in the form which could not be readily available to the plant as in conventional agriculture. The availability of nitrogen is important for the production of phenolic compounds as well as total soluble solids content in plant. Sander and Heitefuss (1998) found in their studies that the increasing nutrient availability to the plant shown decrease in concentration of phenolic compounds. These relationships according to Jones and Hartley (1999) could be explained by different hypothesis, carbon/nutrient balance, growth-differentiation balance and protein competition. It was found by these hypotheses that when the nutrients are more available the plants tend to increase growth and develop more but, on the other hand, the allocation of resources for the production of expendable metabolites like phenolic antioxidants is reduced. In the organic farming system, the slow and prolonged supply of nitrogen does not trigger a spike in plant growth; due to this more photosynthetic sugars could be made available for production of vitamin C and other polyphenols.

### Ascorbic acid content

The vitamin C is among the most important vitamin in fruits and vegetables. Fruit and vegetables including the potato supply more than 90% of the vitamin C in the human diet (Hiza and Bente, 2007). Vitamin C of the horticulture crops could be affected by many pre- and post-harvest factors (Lee and Kader, 2000). Fruit is a better source of antioxidants, flavonoids, phytochemicals and minerals (Ismail and Fun, 2003). Odriozola-Serrano *et al.* (2007) mentioned that vitamin C is an important antioxidant present in fruit and vegetables. According to Hernández *et al.* (2006) vitamin C an additive of processed foods (Rios and Penteado, 2003).

Table 1.4: Ascorbic acid content in mango and pineapple fruits

Fruit	Ascorbic acid content (mg100g <sup>-1</sup> juice)	Origin	Production
Mango cultivar			
Dodo	6.02±1.35bc	Tanzania	Organic
Bolibo	2.36±0.15d	Tanzania	Organic
Viringe	2.46±0.21d	Tanzania	Organic
Tommy Atkins	4.97±0.05c	Burkina Faso	Organic
Kent	8.02±2.60a	Costa Rica	Conventional
Kent	6.93±0.91ab	Ivory Coast	Conventional

Kent	8.40±0.72a	Mali	Conventional
Kent	6.00±0.82bc	Peru	Conventional
Pineapple cultivar			
Smooth cayenne	5.60±1.35bc	Tanzania	Organic
Smooth cayenne	7.47±1.91ab	Uganda	Organic
Smooth cayenne	8.92±0.93a	Costa Rica	Conventional
Smooth cayenne	5.00±1.41c	Ghana	Conventional
Smooth cayenne	6.43±0.67bc	Honduras	Conventional

Values within columns of the same fruits with different letters are significantly different ( $p \leq 0.05$ ).

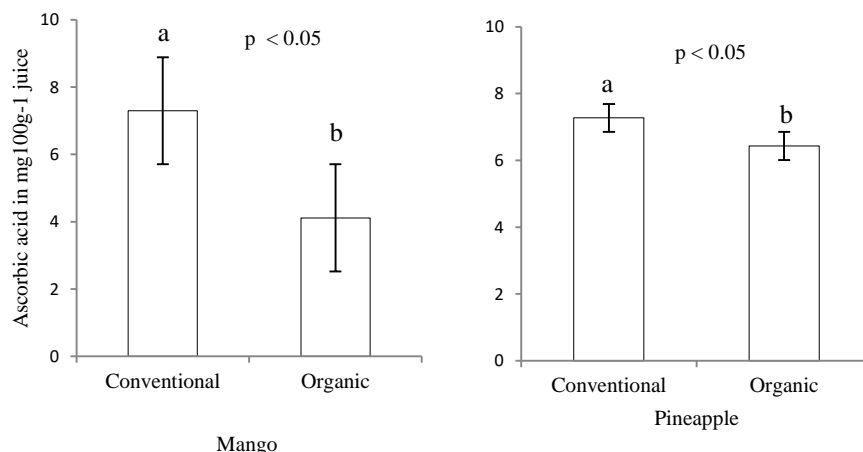


Figure 1.4: Ascorbic acid content in conventional and organic mango (n=12) and pineapple (n=16). Values with different letters are significantly different ( $p \leq 0.05$ ).

In Table 2.4 the ascorbic acid content varied significantly among the mango cultivars. Conventional grown mango have shown on average 43.5% higher ascorbic content than organically grown mango fruits (Fig. 2.4). The levels of ascorbic acid in conventional production system showed that mango cultivar 'Kent' grown in different location varied significantly, suggesting the effect of location due to soil type and climatic condition. The mango cultivars 'Kent' from a conventional production system has shown higher ascorbic content than mango cultivars 'Tommy Atkins', 'Dodo', 'Bolibo' and 'Viringe' from an organic production system. The genetic variability as well as difference in soil type could have an influence on ascorbic acid in mango cultivars

There were significant variations in pineapple cultivars 'Smooth Cayenne' grown in different location under two production systems as indicated in the Table 2.4. Ascorbic acid content was 11.5% higher in conventional than in organic pineapple fruits. The variation of ascorbic acid could be due to the difference in both soil type and production method. According to Virginia (2001), nitrogen from any kind of fertilizer affects the amounts of vitamin C and nitrates as well as the quantity and quality of protein produced by plants. Normally a high supply of nitrogen increases production of protein at the expense of carbohydrates; as a result vitamin C will be reduced. High nitrogen fertilization could be associated with a reduction of ascorbic acid (Lee and Kader, 2000). Wang and Lin (2003) found that the use of compost as a soil supplement has significant effects on ascorbic acid. Organically managed soils generally present plants with lower amounts of nitrogen than chemically fertilized soils, so it would be expected that organic crops would have more vitamin C, less nitrate and less protein, but be of a higher quality than comparable conventional crops. Because our samples were collected from different locations

this might contribute to variation of ascorbic acid, The conventional mango and pineapple fruit have shown higher ascorbic acid levels than organical fruit as indicated in the Fig. 2.4.

## CONCLUSION

The results of this study indicate that differences in cultivar, type of cultivation, location, and climate can significantly influence the levels of ascorbic acid and the total phenolic contents of fruit. Mango cultivars from different locations have shown significant variations in terms of all these variables, while total carotenoids and total phenolic variations were not significant among pineapple cultivars. Organic mango was higher in carotenoids than conventional while total carotenoids content were higher in conventional pineapple were than organic and ascorbic acid was higher in conventional than organic for both mango and pineapple. Antioxidant capacity in both mango and pineapple fruit varied between cultivars, locations and cultivation methods.

## REFERENCES

- Ajila, C.M., K. a. Naidu, S.G. Bhat, and U.J.S.P. Rao. 2007. Bioactive compounds and antioxidant potential of mango peel extract. *Food Chem.* 105(3): 982–988.
- Albrecht, J.. 1993. Ascorbic acid and retention in lettuce. *J. Food Qual.* 16: 311–316.
- Andlauer, W. and Fürst, P. 2002. Nutraceuticals: a piece of history, present status and outlook. *Food Res. Int.* 35: 171–176.
- Anttonen, M. J.; Hoppula, K. I.; Nestby, R.; Verheul, M. J.; Karjalainen, R.O. 2006. Influence of fertilization, mulch color, early forcing, fruit order, planting date, shading, growing environment, and genotype on the contents of selected phenolics in strawberry (*Fragaria × ananassa*Duch.) fruits. *J. Agric. Food Chem* 54: 2614–2620.
- Asami, D.K. , Hong , Y.H. , Barrett , D.M. , Mitchell, A.E. 2003. A comparison of the total phenolic and ascorbic acid contents of freeze-dried and air-dried marionberry, strawberry and corn grown using conventional, organic and sustainable agricultural practices. *J. Agric. Food Chem* 51: 1237 – 1241.
- Balasundram, N., Sundram, K. and Samman, S. 2006. Phenolic compounds in plant and agri-industrial by products: Antioxidant activity, occurrence, and potential uses. *Food Chem.* 1: 191–203.
- Ball, G. 2004. Vitamins: their role in the human body.
- Barrett, D.M., Weakley, C., Diaz, J.V., Watnik, M. 2007. Qualitative and nutritional differences in processing tomatoes grown under commercial organic and conventional production systems. *J. Food Sci* 72: C441 – C450.
- Blomhoff, R., Carlsen, M.H., Andersen, L.F.&Jacobs, D.R.. 2006. Health Benefits of Nuts: Potential Role of Antioxidants. *Br. J. Nutr.*96 (Suppl. 2): 52–60.
- Breithaupt, D.. 2004. Simultaneous HPLC determination of carotenoids used as food colouring additives: applicability of accelerated solvent extraction. *Food Chem.* 86(3): 449–456.
- Cano, P, Plaza, L Sánchez-Moreno, C and de Ancos B. 2003. Elaboración y conservación de zumos de naranja: efectos de nuevastecnologíassobresucalidad sensorial y nutricional, *Alimentación. Nutr. y Salud* 10: 108–119.
- Carbonaro, M., Mattera, M., Nicoli, S., Bergamo, P. and Cappelloni, M. 2002. Modulation of antioxidant compounds in organic vs conventional fruit (peach, *Prunuspersica* L., and pear, *Pyruscommunis* L.). *J. Agric. Food Chem.* 50: pp. 5458–5462.
- Cartea, M.E., M. Francisco, P. Soengas, and P. Velasco. 2011. Phenolic compounds in Brassica vegetables. *Molecules* 16(1): 251–280.

- Chaovanalikit, A. and Wrolstad, R.. 2004. Total anthocyanins and total phenolics of fresh and processed cherries and their antioxidant properties. *Food Chem. Toxicol.* 69(67-72).
- Chassy, A. W.; Bui, L.; Renaud, E. N. C.; Van Horn, M.; Mitchell, A.E. 2006. Three-year comparison of the content of antioxidant micro-constituents and several quality characteristics in organic and conventionally managed tomatoes and bell peppers. *J. Agric. Food Chem* 54: 8244 – 8252.
- Cox, B.D. Whichelow, M.J., and Prevost, A.T. 2000. Seasonal consumption of salad vegetables and fresh fruit in relation to development of cardiovascular disease and cancer. *Publ Heal. Nutr* 3: 19–29.
- Dragsted, L.O., Krath, B., Ravn-haren, G., Vogel, U.B., Vinggaard, A.M., Jensen, P.B., Loft, S., Rasmussen, S.E., Sandstrom, B.M.&Pedersen, A. 2006. Biological Effects of Fruits and Vegetables. *Proc. Nutr. Soc* 65: 61–67.
- Dutta, D., Chaudhuri, U, R, and Chakraborty, R. 2005. Structure, health benefits, antioxidant property and processing and storage of carotenoids. *African J. Biotechnol.* 4(13): 1510–1520.
- Eberhardt, M.V., Lee, C.Y. and Liu, R.H. 2000. Nutrition-Antioxidant activity of fresh apples. *Nature* 405: 903–904.
- Gardner, P.T White, T.A.C. McPhail, D.B. and Duthie, G.G. 2000. The relative contributions of vitamin C, carotenoids and phenolics to the antioxidant potential of fruit juices. *Food Chem.* 68: 471–474.
- Häkkinen, S. H. and Törrönen, A.R. 2000. Content of flavonols and selected phenolic acids in strawberries and *Vaccinium* species: influence of cultivar, cultivation site and technique. *Food Res. Int.* 33: 517–524.
- Halvorsen, B.L., Holte, K., Myhrstad, M.C.W., Barikmo, I., Hvattum, E., Remberg, S.F., Wold, A., Haffner, K., Baugerod, H., Andersen, L.F., Moskhaug, J.Ø., Jacobs, D.R.JR&Blomhoff, R. 2002. A Systematic Screening of Total Antioxidants In Dietary Plants. *J. Nutr* 132: 461–471.
- Hassimoto, N.M.A, Genovese M.I and Lajolo, F.M. 2005. Antioxidant activity of dietary fruits, vegetables, and commercial frozen fruit pulps. *J. Agric. Food Chem.* 53: 2928–2935.
- Joomwong, A.. 2006. Impact of cropping season in northern Thailand on the Quality of smooth cayenne pineapple. II. Influence on Physico-chemical attributes. *Int. J. Agri. Biol.* 8(3): 330 – 336.
- Kauer C, K.H.C. 2001. Antioxidants in fruits and vegetables—the millenium’s health. *Int J. Food SciTechnol* 36: 703–725.
- Kris-Etherton, P.M., Hecker, K.D., Bonamone, A., Coval, S.M., Binkoski, A.E., Hilpert, K.F., Griel, A.E., Etherton, T.D. 2002. Bioactive compounds in foods: their role in the prevention of cardiovascular disease and cancer. *Am J Med* 113: 71–81.
- Lattanzio, V. 2003. Bioactive polyphenols: their role in quality and storability of fruit and vegetables. *J. Appl. Bot.* 77: 128–146.
- Lombardi-Boccia, G. Lucarini, M. Lanzi, S. Aguzzi, A. and C. 2004. Nutrients and Antioxidant Molecules in Yellow Plums (*Prunus domestica* L.) from Conventional and Organic Productions: A Comparative Study. *J. Agric. Food Chem* 52: 90–94.
- Lundegårdh, B. 2005. Organic cultivation systems and food quality. NJF-Seminar 369. Organic farming for a new millenium - status and future challenges Nordiskajordbruksforskarensfrening (NJF).
- Mortensen, A. 2006. Carotenoid and other pigments as natural colourants. *Pure Appl. Chem* 78(8): 1477–1491.
- Moure A., Cruz J.M., Franco D., Dominguez J.M., Sineiro J., Dominguez H., Nunez M.J., Parajo J.C. 2001. Natural antioxidants from residual sources. *Food Chem.* 72: 145–171.

- Oberholtz, L., Dimitri, C., Greene, C. 2005. Price Premiums Hold on as U.S. Organic Produce Market Expands; Outlook Report VGS30801; U.S. Department of Agriculture, Economic Research Service: Washington, DC.
- Ohr, L. 2004. Dietary antioxidants. *Food Technol.* 58(10): 67–74.
- Podsedeck, A. 2005. Natural antioxidants and antioxidant capacity of Brassica vegetables. *A Rev. LWT* 40: 1–11.
- Rao, A.V. and Rao, L.G. 2007. Carotenoids and human health. *Pharmacol. Res* 55: 207 – 216.
- Ribeiro, S.M.R., L.C. a Barbosa, J.H. Queiroz, M. Knödler, and a. Schieber. 2008. Phenolic compounds and antioxidant capacity of Brazilian mango (*Mangifera indica* L.) varieties. *Food Chem.* 110(3): 620–626.
- Ribeiro, S.M.R, de Queiroz, J.H, de Queiroz, M.E.L.R, Campos, F.M and Sant’ana, H.M.. 2007. Antioxidant in mango(*mangiferaIndica* L.) pulp. *Plant Food Hum. Nutr.* 62: 13–17.
- Rodriguez-Amaya D.B. 2001. A guide to Carotenoid Analysis in Foods. ILSI Press, Washington DC, USA.
- Sánchez-Moreno, C., Plaza, L de Ancos, B, and Cano, P. 2006. Nutritional characterization of commercial traditional pasteurized tomato juices: carotenoids, vitamin C and radical-scavenging capacity. *Food Chem.* (98): 749–756.
- Sandmann, G. 2001. Genetic manipulation of carotenoid biosynthesis, strategies, problems and achievements. *Trends Plant Sci* 6: 14 – 17.
- Shepherd, R., Magnusson, M., Sjoden, P.O. 2005. Determinants of consumer behaviour related to organic foods. *Ambio* 34: 352–359.
- Singleton, V.L. and Rossi, J.A. 1965. Colorimetry of total phenols with phosphomolybdic-phosphotungstic reagents. *Am. J. Enol. Vitic* 16: 144–158.
- Stracke, B.A., Rufer, C.E., Weibel, F.P., Bub, A., Watzl, B. 2009. Three-year comparison of the polyphenol contents and antioxidant capacities in organically and conventionally produced apples (*Malus domestica* Bork. cultivar “Golden Delicious.” *J. Agric. Food Chem.* 57: 4598–4605.
- Szeto, Y.T., Brian, T. and Benzie, I.F.. 2002. Total antioxidant and ascorbic acid content of fresh fruits and vegetables: implications for dietary planning and food preservation. *Br J Nutr* 87: 55–59.
- Trombino, S., Serini, S., Di Nicuolo, F., Celleno, L., Ando, S., Picci, N., Calviello, G. and Palozza, P. 2004. Antioxidant effect of ferulic acid in isolated membranes and intact cells: Synergistic interactions with  $\beta$ -tocopherol,  $\beta$ -carotene and ascorbic acid. *J. Agric. Food Chem.* 52: 2411–2420.
- Usenik, V., Mikulič-Petkovšek, M., Solar, A. and Ätampar, F. 2004. Flavonols of leaves in relation to apple scab resistance. *Zeitschrift für Pflanzenkrankheiten und Pflanzenschutz* 111: 137–144.
- Valavanidis, A., Vlachogianni, T., Psomas, A., Zovoili, A., Siatis, V. 2009. Polyphenolic profile and antioxidant activity of five apple cultivars grown under organic and conventional agricultural practices. *Int’l J. Food Sci. Tech* 44: 1167–1175.
- Veberic, R., and and F. Stampar. 2005. Quality of Apple Fruits ( *Malus domestica* ) from Organic Versus Integrated Production. FRUTIC.
- Weibel, F., Bickel, R., Leuthold, S., Alfoldi, T. 2000. Are organically grown apples tastier and healthier? A comparative field study using conventional and alternative methods to measure fruit quality. *Acta Hort* 57: 417 – 426.
- Wellburn, A.R. 1994. The spectral determination of chlorophyll a and b as well total carotenoid, using various solvents with Spectrophotometers of different resolutions. *J. Plant Physiol* 144: 307–313.

Winter, C.K. and Davis, C.F. 2006. Organic foods. *J. Food Sci* 71: 117 – 124.

Young, J.E.; Zhao, X.; Carey, E. E.; Welti, R., Yang, S.S., Wang, W. 2005. Phytochemical phenolics in organically grown vegetables. *Mol. Nutr. Food Res* 49: 1136 1142.

Zulueta, A., M.J. Esteve, I. Frasquet, and A. Frígola. 2007. Vitamin C, vitamin A, phenolic compounds and total antioxidant capacity of new fruit juice and skim milk mixture beverages marketed in Spain. *Food Chem.* 103(4): 1365–1374.

Abbreviations-AA = Ascorbic acid, GAE = Gallic acid equivalents

FW = Fresh weight