Vol.1, No.1, pp.12-16, September 2016

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REMOTE ISCHEMIC PRECONDITIONING PROTECTS AGAINST ISCHEMIC REPERFUSION INJURY DURING HEART TRANSPLANTATION

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ABSTRACT: Background: Remote ischemic pre- conditioning (RIPC) is a technique where the application of short, repetitive limb intervals of ischemia and reperfusion would result in the condition of the heart as well as other organs to tolerate the ischemic reperfusion injury. Methods: Many research groups are interested in investigating the mediators, through which this technique works. Many mediators have been suggested to mediate the protective actions of RIPC.Conclusion: This manuscript represents a short review explaining a possible mechanism of action of this technique, through recent published findings, which confirm the ability of the technique to play an important role in the clinical practice of heart transplantation.

KEYWORDS: Remote Ischemic Preconditioning, Ischemic Preconditioning, Nitric Oxide, Inflammasomes, Inflammatory Cytokines, And Heart Transplantation.

INTRODUCTION

Ischemic preconditioning is a technique where prior application of repeated short cycles of ischemia and reperfusion would be able to attenuate the severity of subsequent ischemic reperfusion injury (IRI). Remote ischemic preconditioning (RIPC) describes the ability of the technique to function through distance. For example, the application of short, repetitive ischemia- reperfusion cycles of the limb would protect distant organs like heart, kidney, brain and liver during subsequent IRI. Both phenomena indicate the involvement of local, paracrine as well as remote, circulating mediators (1).

During limb ischemia, the diminished flow and shearing stress would be associated with cell membrane depolarization and inhibition of the inward driving K⁺ channels. The inhibition of KATP channels would lead to the activation of T type Ca^{2+} channels and increased Ca^{2+} influx into endothelial cells. Increased intracellular Ca^{2+} activates Ca^{2+} - dependent endothelial NO synthase (eNOS) (1,2).

Simultaneously, hypoxia and ischemia would result in an increased production of reactive oxygen species (ROS). Hypoxia inhibits oxidative phosphorylation and results in decreased ATP production. Which augments the inhibition of KATP channels and activates xanthine oxidase, leading to increased ROS production. Inhibition of KATP channels and cell membrane depolarization would also result in NADPH oxidase (NOX2) activation, leading to more increase of ROS production (1,3).

Increased production of both NO and ROS might be associated with NO oxidation to produce nitrite (NO_2^{-}) . Several studies documented the important role of NO in mediating the protective effect of IPC and RIPC. While the locally produced NO can exert its action in case of IPC, it can't be accused for RIPC protective effect because of its short blood half-life (≤ 2 milliseconds) (4). However, it was observed that NO inhalation in human provides protection against IRIs, while being associated with a significant increase in the circulating levels of

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nitrite. In addition, NO_2^- showed the ability to protect against IRI, to exert cytoprotective effects, and to decrease the infarction size similar to NO (5-12). Moreover, it has recently been confirmed that the application of brachial artery RIPC results in the activation of eNOS and increased plasma NO_2^- levels (13).

In the heart, NO_2^- would be reduced to NO and N_2O_3 by myoglobin (14,15). NO and Snitrosothiols formed from nitrite would inhibit complex I of the respiratory chain during reperfusion. This would attenuate the increased production of ROS in response to IRI, and would indirectly affect the functionality of complex II (16,17). Being at cross-talking with mitochondrial KATP channels, modification of the functional activity of complex II would influence the activity of mitochondrial KATP channels (18), this might contribute to an improved activity of these channels in response to RIPC, which would inhibit the opening of mitochondrial permeability transition pores and the subsequent release of cytochrome c during reperfusion (17,19).

An important mechanism in the development of the IRI would be the increased production of inflammatory cytokines, which would be responsible for the recruitment of inflammatory cells and initiation of adverse inflammatory reactions (20). In addition to the significant increase of ROS production, IRI activates toll- like receptors (TLRs). Both result in priming of the heart inflammasomes (21).

During ischemia and hypoxia, as well as cold preservation of the heart graft, the associated inhibition of Na⁺- K⁺ ATPase and or other K⁺ channels would result in decreased intracellular K⁺ levels. Even with the administration of high extracellular K⁺ concentrations (during cardioplegia), this would lead to the closure of K⁺ channels (3). The end result would be the drop of intracellular K⁺ levels, which activates the primed inflammasomes (22).

Activated inflammasomes activate caspase- 1, which activates proIL1 β and proIL18. With IL1 β being able to induce IL6, and with a confirmed important role of inflammasomes and TLRs in the establishment of inflammatory reactions and IRI, the above described role of NO and NO₂⁻ to attenuate ROS production and to improve the activity of KATP channels would interfere with inflammasomes priming and activation in response to IRI. Accordingly, this would contribute to decreased production of inflammatory cytokines, which would ultimately attenuate the immune cell infiltration and the adverse immune reactions generated in response to IRI. (Figure 1)

This mechanism of action of RIPC highlights the importance of inflammasomes inactivation to attenuate the hazards of IRI, though it was reported to the contrary by some studies that the deletion of NLRP3 (which is the most studied inflammasome component) abates the protective effects of IPC due to inhibition of IL6 production and lacking of its signaling (23). It seems that various inflammatory cytokines are involved in the stimulation of the adverse inflammatory reactions in response to IRI, as well as in protective signaling against subsequent IRI. Accordingly, the above discussed scenario should be confirmed as a whole by experimental studies, to identify whether blocking the release of IL1 β and IL18, with the subsequent lack of IL6 induction, would increase or decrease heart protection.

Nevertheless, the augmentation of the above presented scenario at different levels (for example, through NO inhalation, NO_2^- administration, the use of KATP channel agonists) prior to heart transplantation and or other cardiac IRIs was found to (or is expected to)

International Journal of Nutrition and Metabolism Research

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provide a significant degree of protection against the adverse effects of IRI, with an associated better clinical outcome (24).

In conclusion, this manuscript reviews a possible mechanism, through which RIPC would be involved in the attenuation of inflammasomes activation and cytokine production within the heart in response to IRI, especially during transplantation. Further studies should be conducted to confirm this mechanism, and whether it could be also considered for other organs such as lung, kidney and liver.

Conflicts of interest

No special funding was provided to produce this work. The author declares no conflicts of interest.

REFERENCES

1. Paola Corti and Mark T. Gladwin. Is Nitrite the Circulating Endocrine Effector of Remote Ischemic Preconditioning? Circ Res. 2014;114:1554-1557.

2. Wei Z, Manevich Y, Al-Mehdi AB, Chatterjee S, and Fisher AB. Ca2+ flux through voltage-gated channels with flow cessation in pulmonary microvascular endothelial cells. Microcirculation 11: 517-526, 2004.

3. Shampa Chatterjee, Gary F. Nieman, Jason D. Christie and Aron B. Fisher. Shear stress-related mechanosignaling with lung ischemia: lessons from basic research can inform lung transplantation. Articles in PresS. Am J Physiol Lung Cell Mol Physiol (September 19, 2014)

4. Liu X, Miller MJ, Joshi MS, Sadowska-Krowicka H, Clark DA, and Lancaster JR Jr. Diffusion-limited reaction of free nitric oxide with erythrocytes. J Biol Chem. 1998;273:18709–18713.

5. Gladwin MT, Ognibene FP, Pannell LK, et al. Relative role of heme nitrosylation and beta-cysteine 93 nitrosation in the transport and metabolism of nitric oxide by hemoglobin in the human circulation. Proc Natl Acad Sci U S A. 2000;97:9943–9948.

6. Cannon RO 3rd, Schechter AN, Panza JA, et al. Effects of inhaled nitric oxide on regional blood flow are consistent with intravascular nitric oxide delivery. J Clin Invest. 2001;108:279–287.

7. Gladwin MT, Shelhamer JH, Schechter AN, et al. Role of circulating nitrite and Snitrosohemoglobin in the regulation of regional blood flow in humans. Proc Natl Acad Sci U S A. 2000;97:11482–11487.

8. Cosby K, Partovi KS, Crawford JH, et al. Nitrite reduction to nitric oxide by deoxyhemoglobin vasodilates the human circulation. Nat Med. 2003;9:1498–1505.

9. Webb A, Bond R, McLean P, Uppal R, Benjamin N, Ahluwalia A. Reduction of nitrite to nitric oxide during ischemia protects against myocardial ischemia-reperfusion damage. Proc Natl Acad Sci U S A. 2004;101:13683–13688.

10. Duranski MR, Greer JJ, Dejam A, et al. Cytoprotective effects of nitrite during in vivo ischemia-reperfusion of the heart and liver. J Clin Invest. 2005;115:1232–1240.

11. Gonzalez FM, Shiva S, Vincent PS, et al. Nitrite anion provides potent cytoprotective and antiapoptotic effects as adjunctive therapy to reperfusion for acute myocardial infarction. Circulation. 2008;117:2986–2994.

12. Neye N, Enigk F, Shiva S, et al. Inhalation of NO during myocardial ischemia reduces infarct size and improves cardiac function. Intensive Care Med. 2012;38:1381–1391.

Vol.1, No.1, pp.12-16, September 2016

Published by European Centre for Research Training and Development UK (www.eajournals.org)

13. Rassaf T, Totzeck M, Hendgen-Cotta UB, Shiva S, Heusch G, Kelm M. Circulating nitrite contributes to cardioprotection by remote ischemic preconditioning. Circ Res. 2014;114:1601–1610.

14. Sparacino-Watkins CE, Tejero J, Sun B, et al. Nitrite reductase and nitric-oxide synthase activity of the mitochondrial molybdopterin enzymes mARC1 and mARC2. J Biol Chem. 2014;289:10345–10358.

15. Tejero J, Gladwin MT. The globin superfamily: functions in nitric oxide formation and decay. Biol Chem. 2014 Jun;395(6):631-9.

16. Shiva S, Sack MN, Greer JJ, et al. Nitrite augments tolerance to ischemia/reperfusion injury via the modulation of mitochondrial electron transfer. J Exp Med. 2007;204:2089–2102.

17. Chouchani ET, Methner C, Nadtochiy SM, et al. Cardioprotection by S-nitrosation of a cysteine switch on mitochondrial complex I. Nat Med. 2013;19:753–759.

18. Testai L, Rapposelli S, Martelli A, Breschi MC, Calderone V. Mitochondrial Potassium Channels as Pharmacological Target for Cardioprotective Drugs. Med Res Rev. 2014 Oct 26. [Epub ahead of print]

19. Shiva S, Huang Z, Grubina R, et al. Deoxymyoglobin is a nitrite reductase that generates nitric oxide and regulates mitochondrial respiration. Circ Res. 2007;100:654–661.

20. Kawaguchi M, Takahashi M, Hata T, et al. Inflammasome activation of cardiac fibroblasts is essential for myocardial ischemia/reperfusion injury. Circulation 2011; 123(6): 594-604.

21. Fernandes-Alnemri T, S Kang, C Anderson, J Sagara, K A Fitzgerald, and E S Alnemri. Cutting edge: TLR signaling licenses IRAK1 for rapid activation of the NLRP3 inflammasome. J. Immunol 2013; 191: 3995–3999.

22. Mohammed G. Ghonime, Obada R. Shamaa, Srabani Das, et al. Inflammasome Priming by Lipopolysaccharide is Dependent upon ERK Signaling and Proteosome Function. J Immunol 2014; 192: 3881- 3888.

23. Coert J. Zuurbier, Willeke M. C. Jong, Otto Eerbeek, Anneke Koeman, Wilco P. Pulskens, Loes M. et al. Deletion of the Innate Immune NLRP3 Receptor Abolishes Cardiac Ischemic Preconditioning and Is Associated with Decreased II-6/STAT3 Signaling. PLoS ONE 2012; 7 (7): e40643.

24. Sivaraman V, Yellon DM. Pharmacologic therapy that simulates conditioning for cardiac ischemic/reperfusion injury. J Cardiovasc Pharmacol Ther. 2014 Jan;19(1):83-96.

International Journal of Nutrition and Metabolism Research

Vol.1, No.1, pp.12-16, September 2016

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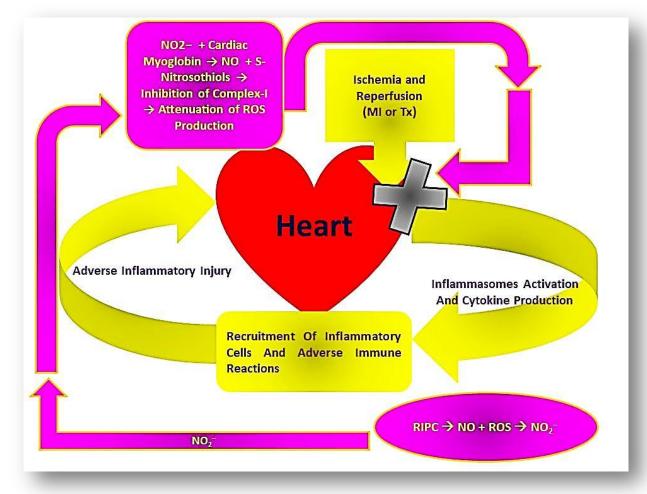


Figure 1; diagrammatic representation of the mechanism, through which RIPC attenuates inflammasomes activation and cytokine production within the heart in response to IRI.