Published by European Centre for Research Training and Development UK (www.eajournals.org)

INDUCTION OF EPITHELIAL CELL APOPTOSIS BY ACINETOBACTER BAUMANNII A424

*, **Lubna Alzubaidi, ***Cordula Stover

*University of Al Mustansiria, College of Engineering, Department of Environmental Engineering, Baghdad Iraq **Postdoctoral researcher at the University of Leicester (UK) **University of Leicester, College of Medicine, Department of infection, immunity and inflammation (UK)

ABSTRACT: Acinetobacter baumannii is considered as highly contagious pathogen, it can cause serious infections in the Skin, lungs, blood, and brain. It may also cause burn, urinary tract and wound infections. It can be spread by direct contact and may be found on skin or in food, water, or soil. It may also be found in hospitals. Learning how dose A.baumannii effect on cells and program of cell death has been a major research goal for researchers. In current study, we chose Acinetobacter baumannii A424 strain as a model for cell apoptosis since little information are available for this strain. Live bacteria were more likely to produce epithelial cell death after few hours of incubation. Killed and filtrate form of this strain gave moderate apoptotic activity in HACAT cell, degraded DNA of apoptotic cell showed approximately 140 pb in size. In conclusion the ability of A424 strain of Acinetobacter baumannii to cause cell apoptosis as well as other o9ther characteristics like multidrug resistance activity, biofilm formation and survival in desiccation (data not shown) may reflect the virulence characteristics of this opportunistic strain which may infect ill or severely wounded patients.

KEYWORDS: Acinatobacter baumannii, Cell apoptosis, virulence factor.

INTRODUCTON

In recent years *Acinetobacter baumannii* has increasingly been responsible for human nosocomial infections. Acinetobacter is a killer bug in some departments in some hospitals. As a hospital pathogen, *A. baumannii* mainly affects patients in the intensive care unit (ICU), including burn patients, trauma patients, and patients requiring mechanical ventilation. Also, any immunocompromised patient or anyone who has an underlying disease, such as chronic lung disease or diabetes, is at an increased risk for *A. baumannii* infection. These bacteria were nomnanted recently as "Iraqibacter" because of its origin in military hospitals in Iraq and its persistence among veterans. (1,2,3) Interest in Acinetobacter, from both the scientific and public community, has risen sharply over recent years.(4,5,6) .The full genome sequencing of *Acinetobacter baumannii* shows that this organism harbors a remarkable number of putative virulence-associated genes and elements homologous to the Legionella/Coxiella type IV secretion apparatus(7,8). Several virulence determinants, such as biofilm formation, adherence and ability

European Journal of Biology and Medical Science Research

Vol.5, No.2, pp.20-26, April 2017

Published by European Centre for Research Training and Development UK (www.eajournals.org)

to invade host cells were detected in *A.baumannii* (9). Some infections associated with *A.baumannii* include ventilator-associated pneumonia, skin and soft-tissue infections, secondary meningitis, urinary tract infections, wound and blood stream infections, endocarditis, intraabdominal abscess, and surgical site infections.10. In this study we the ability of *A.baumannii* A424 to cause apoptosis in HACAT epithelial cells.

MATERIALS AND METHODS

Bacterial strain and growth conditions

Acinetobacter baumannii A424 strain was obtained from Dr.K.Rajakumar University of Leicester (UK). Prior each experiment, a single colony of baumannii A424 was inoculated into 5ml Luria broth and grown at 37C with shaking for 18 hour. Bacterial culture was washed three times with buffer saline (PBS at PH 7.2) inoculated to cell line culture (whole cell), killed bacteria were also used by exposing bacteria to 60C for 15 min. filtrate culture was prepared by inoculation *A.baumannii A424* to LB medium, the cultured bacteria were centrifuged and the supernatants were filtrated using 0.22µm. references

Cell line culture

Human HACAT epithelial cell was grown in DMEM medium supplemented with 10% fetal calf serum (FCS), 100U of penicillin per ml and 100mg streptomycin per ml at 37C in 5%CO2. Cells were observed daily under light microscope.

Induction of cell apoptosis

HACAT epithelial cells were treated with *A.baumannii* A424 (whole cells, killed bacteria and filterate). Cells were observed after (2, 4, 8, 16, 24 and 48h). 10⁸ CFU were used for whole bacteria Reference. Infected cells were incubated at 37C with 5%CO2.

Detection of cell death

a.Trypan blue uptaking: After 24h of incubation, cells apoptosis were examined for using trypan blue stain. Cells were washed gently three times with PBS, and stained with 0.4% for 5 min. cells were examined by inverted phase contrast light microscopy and cells count were detected using hemocyotometer. (11)

b.DNA fragmentation was carried out according to the protocol of (12) with slight modification, DNA from The adherent HACAT cells were harvested and analyzed for DNA fragmentation. Cells were collected by scrapping, washed in PBS and treated with Triton X100 (source or company), cells were collected and centrifuged at 3500rpm for 15min. cells were re-suspended and treated with RNase for 37C for 1 hour. 20 μ g/ml of protease K was added (50C for 2 hours). DNA extraction was extracted with phenol, chloroform, isoamyle alcohol (25, 24, 1), and centrifuged, to precipitate DNA, supernatant was treated with equal volume of isopropanol with overnight Published by European Centre for Research Training and Development UK (www.eajournals.org) incubation at -20C. Sample was centrifuged (4C, 15min) and treated with TBA (Tris boric acid), analysis was done on a 1.5% agarose gel.

Electron microscopy: Infected HACAT epithelial cells were grown in 35mm culture dishes at concentration of 10^6 cells/ well. *A baumannii* A424 was infected to HACAT cells with MOI 100, 50 and incubated for 18h (7), the cells were washed four times with PBS and harvested by using tryton X100 and re-suspended in appropriate fixative. Samples were sent to the electron microscopy section at the University of Leicester for further investigation.

RESULTS

Learning how *A.bamannii* effect on cells and program of cell death has been a main goal for researcher, In the current study we chose *A.baumannii* A424 as a model for this bacterium and according to the knowledge of Author, no information are available about cell apoptosis by this strain of Acinetobacter.

Induction of HACAT cell apoptosis:

To investigate the ability A424 of Acinetobacter strain to invade HACAT epithelial cells, whole cell, killed and filtrate form was applied. (Fig 1), whole cell caused noticeable apoptotic activity after 24h and 90% apoptosis was detected after 48h., killed and filtrate bacteria had less apoptotic activity after 24h than whole bacteria. rther processing.



Fig.1. Effect of live bacteria killed and filtrate on viability of HACAT Epithelial cell. Cells were co-cultured with *Acinetobacter baumannii* A424 (2,4,8,16,24,48h)

Published by European Centre for Research Training and Development UK (www.eajournals.org)



A.Healthy cell B. after 24 hour C. After 48hour

Fig.2. A. illustrates the morphology of healthy HACAT epithelial cell, B,C infected cells after 24, 48 h



Fig.3. A trypan blue positive cells after 24h B .control cells.



Fig.4. DNA fragmentation of cell after infection with A.baumannii A424

Published by European Centre for Research Training and Development UK (www.eajournals.org)



Fig.5.Electron microscopy for Acinetobacter baumannii A424

DISCUSSION

Multidrug-resistant Acinetobacter baumannii is a formidable threat and associated with wide spectrum of infectious diseases including nosocomial infections, community-acquired and those following war or natural disaster especially to military personnel with war wound. The treatment has become difficult, not only because the bacterium can develop extensive antimicrobial resistance but because it also forms biofilms that are resistant to host defense as well as causing host cell apoptosis.(13,2) .In current study Acinetobacter baumannii A424 strain was chosen as a model to see the ability of this strain to induce HACAT cells apoptosis, data illustrated that whole bacteria had a strong apoptotic activity after 24h, less among killed and filtrate form, this indicates that living cell of A424 strain may contain apoptosis including molecule(s), that may activate the program of cell death and increases the virulence ability of this bacteria, Although other virulence factors were confirm for this strain including biofilm formation, survival in desiccation and multidrug resistant activity (data not shown), it has been reported that the apoptosis of epithelial cells was induced by various components of bacteria. (14). Activation of casebase is an important step for cell apoptosis, investigations demonstrated that the precursor form of casepase 3 is cleaved by live A.baumannii to produce active form by live bacteria (5). The ability of bacteria to induce host cell apoptosis is considered to be an important bacterial virulence mechanism.(3). Bacteria may even up-regulate the endogenous receptor/ligand system that induces apoptosis, generally when the bacteria are bound to the host cell.(15), it was noticed in current study that HACAT apoptosis was started at early stage for whole lived bacteria. A study carried out by (16-18) demonstrated that Omp38 is considered as potent cytotoxin that induces apoptosis of epithelial cells in A.baumannii and this may explain the strong effect of live bacteria on cell apoptosis. In current study cell apoptosis was also detected with filterate form of A424 baumannii this finding suggests that this strain may produce certain products that mediate cell apoptosis. (19,5).

In conclusion the ability of A424 strain of *Acinetobacter baumannii* to cause cell apoptosis as well as other characteristics like multidrug resistance activity, biofilm formation and survival in

Published by European Centre for Research Training and Development UK (www.eajournals.org)

desiccation (data not shown) may reflect the virulence characteristics of this opportunistic strain which may infect ill or severely wounded patients.

REFERENCES

- Ian P. Mumm,a Thammajun L. Wood,b Karthik R. Chamakura,a Gabriel F. Kuty Everetta. Complete Genome of Acinetobacter baumannii Podophage Petty.Genome A. 2013 Volume 1 Issue 6.
- Dallo SF, Weitao T. Insights into Acinetobacter.war-wound infections, biofilms, and control. Adv Skin Wound Care. 2010 Apr; 23(4):169-74.
- Marcelo Lancellotti1, Rafaella Fabiana Carneiro Pereira1, Gisele Gentile Cury1 and Luciana Maria de Hollanda. Pathogenic and Opportunistic Respiratory Bacteria-Induced Apoptosis. The Brazilian Journal of Infectious Diseases and Contexto. 2009;13(3):226-231
- Anton Y. Peleg, Harald Seifert and David L. Paterson. 2008. Acinetobacter baumannii: Emergence of a Successful Pathogen. American Society for Microbiology. Vol. 21, No. 3.
- LEE, J. Y. OH, K. S. KIM, Y. W. JEONG, J. C. PARK and J. W. CHO. 2001. Apoptotic cell death induced by Acinetobacter baumannii in epithelial cells through caspase-3 activation. APMIS 109: 679–84
- Callie Camp, MS, MT(ASCP)CM, Owatha L. A Review of Acinetobacter baumannii as a Highly Successful Pathogen in Times of War.2010. Lab Med. 41(11):649-657.
- Jong Sook Jin, Sang-Oh Kwon2, Dong Chan Moon1, Mamata Gurung1, Jung Hwa Lee1, Seung Il Kim2, Je Chul Lee 2011. Acinetobacter baumannii Secretes Cytotoxic Outer Membrane Protein A via Outer Membrane Vesicles PLoS ONE. Volume 6 | Issue 2 | e17027.
- Michael G. Smith, Tara A. Gianoulis, Stefan Pukatzki, John J. Mekalanos, L. Nicholas Ornston, Mark Gerstein, and Michael Snyder. 2007. New insights into Acinetobacter baumannii pathogenesis revealed by high-density pyrosequencing and transposon mutagenesis GENES & DEVELOPMENT 21:601–614
- Jin Yeol Park, Shukho Kim, Sung-Min Kim, Sun Ho Cha, Si-Kyu Lim,and Jungmin. 2011. Complete Genome Sequence of Multidrug-Resistant Acinetobacter baumannii Strain 1656-2, Which Forms Sturdy Biofilm. American Society for Microbiology. Vol. 193, No. 22.
- Jeremy A. Iwashkiw1, Andrea Seper2, Brent S. Weber1, Nichollas E. Scott1,3, Evgeny Vinogradov4, Chad Stratilo5, Bela Reiz6, Stuart J. Cordwell3, Randy Whittal6, Stefan Schild2, Mario F. Feldman. Identification of a General O-linked Protein Glycosylation System in Acinetobacter baumannii and Its Role in Virulence and Biofilm Formation. June 2012 | Volume 8 | Issue 6 | e1002758.
- Crane JK, Majumdar S, Pickhard III, Host cell death due to enteropayhogenic Escherichia coli has features of apoptosis. Infect Immun. 1999;67:2575-84
- Chul Hee Choi, Eun Young Lee, Yoo Chul Lee, Tae In Park, Hwa Jung Kim, Sung Hee Hyun, Soon Ae Kim, Seong-Kyu Lee and Je Chul Lee. Outer membrane protein 38 of Acinetobacter baumannii localizes to the mitochondria and induces apoptosis of epithelial cells. Cellular Microbiology (2005) 7(8), 1127–1138.
- Characterizationof the Acinetobacter baumannii growth phase-dependent andserumresponsive transcriptomes Anna C. Jacobs1,2, Khalid Sayood3, Stephen B. Olmsted2, Catlyn E. Blanchard2, Steven Hinrichs1, David Russell3 & Paul M. Dunman FEMS Immunol Med Microbiol 64 (2012) 403–412.

```
Vol.5, No.2, pp.20-26, April 2017
```

Published by European Centre for Research Training and Development UK (www.eajournals.org)

- Moss JE, Idanpann-Heikkila I, Zychlinsky A. Induction of apoptosis by microbial pathogens. Cellular microbiology. Washington DC:ASM press, 2000:275-90.
- Joly-Guillou, Burn- Buisson. Epidemiology of Acinetobacter spp: surveillance and manaemnt of outbreak.Microbiology,Epidemiology, infections, management. Florida: CRC press, 1996:71-100.
- Choi CH, Lee EY, Lee YC. Outer membrane protein 38 of Acinetobacter baumannii localizes to mitochondria and induces poptosis of epithelial cells. Cell Microbiol 7:1127-1138.
- Identification of a General O-linked Protein Glycosylation System in Acinetobacter baumannii and Its Role in Virulence and Biofilm Formation. Jeremy A. Iwashkiw1, Andrea Seper2, Brent S. Weber1, Nichollas E. Scott1,3, Evgeny Vinogradov4, Chad Stratilo5, Bela Reiz6, Stuart J. Cordwell3, Randy Whittal6, Stefan Schild2, Mario F. Feldman1. June 2012 | Volume 8 | Issue 6 | e1002758. PLoS Pathogens.
- Anton Y. Peleg,1* Harald Seifert,2 and David L. Paterson. Acinetobacter baumannii: Emergence of a Successful Pathogen. CLINICAL MICROBIOLOGY REVIEWS, July 2008, p. 538– 582. Vol. 21, No. 3
- Leung WS, Chu CM, Tsang KY, Lo FH, Lo KF, Ho PL. Fulminant community-acquired Acinetobacter baumannii pneumonia as a distinct clinical syndrome. Chest. 2006 Jan; 129(1):102-9.
- Hanuszkiewicz A, H€ubner G, Vinogradov E, Lindner B, Brade L, Brade H, Debarry J, Heine H, Holst O (2008):Structural and immunochemical analysis of the lipopolysaccharide from Acinetobacter lwoffii F78 located outside Chlamydiaceae with a Chlamydia-specific lipopolysaccharide epitope. Chem Eur J 14:10251–10258

AKNOWLEDEMENTS

This study was supported by Iraqi Ministry of higher education and scientific research As postdoctoral research followship. The Author thank Dr.K.Rajakumar who generously provided Acinetobacter baumannii A424 strain, the author also thank the staff in electron microscope in the University of Leicester (UK) for their help and assistance.